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Biflavonoids. Part 4^{1*}. Structure and Stereochemistry of Novel Flavanone- and the First Isoflavanone-benzofuranone Biflavonoids

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Abstract: The structure and stereochemistry of (2*R*,3*S*)-naringenin-(3 α →5)-(2*R*)-maesopsin, its isoflavanone analogue, (2*S*,3*R*)-dihydrogenistein-(2 α →7)-(2*R*)-maesopsin, and the 2*S* (F-ring) diastereomers of both compounds from the heartwood of *Berchemia zeyheri* were established by ¹H NMR and CD data. The isoflavanone analogues represent the first isoflavanone-benzofuranone biflavonoids.

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INTRODUCTION

Our first reports^{2,3} on the phenolic constituents of the red heartwood of *Berchemia zeyheri* revealed the presence of a flavanone-benzofuranoid type dimer, (2*R*,3*S*)-naringenin-(3 α →7)-(2*R*)-maesopsin (zeyherin)⁴ and its 2-C(F) diastereomer, (2*R*,3*S*)-naringenin-(3 α →7)-(2*S*)-maesopsin. The collective utilization of chemical degradation, ¹H NMR, CD and computational data not only defined the structures but permitted estimation of the absolute stereochemistry of the zeyherin diastereomers as well as related structures from the same source.^{2,3}

Continued investigation has now revealed the existence of a second diastereomeric pair of regio-isomers, (2*R*,3*S*)-naringenin-(3 α →5)-(2*R*)-maesopsin **1** and (2*R*,3*S*)-naringenin-(3 α →5)-(2*S*)-maesopsin **2** as well as a unique pair of related diastereomers, (2*S*,3*R*)-dihydrogenistein-(2 α →7)-(2*R*)-maesopsin **5** and (2*S*,3*R*)-dihydrogenistein-(2 α →7)-(2*S*)-maesopsin **6**, representing the first natural isoflavanone-benzofuranoid oligomers.¹

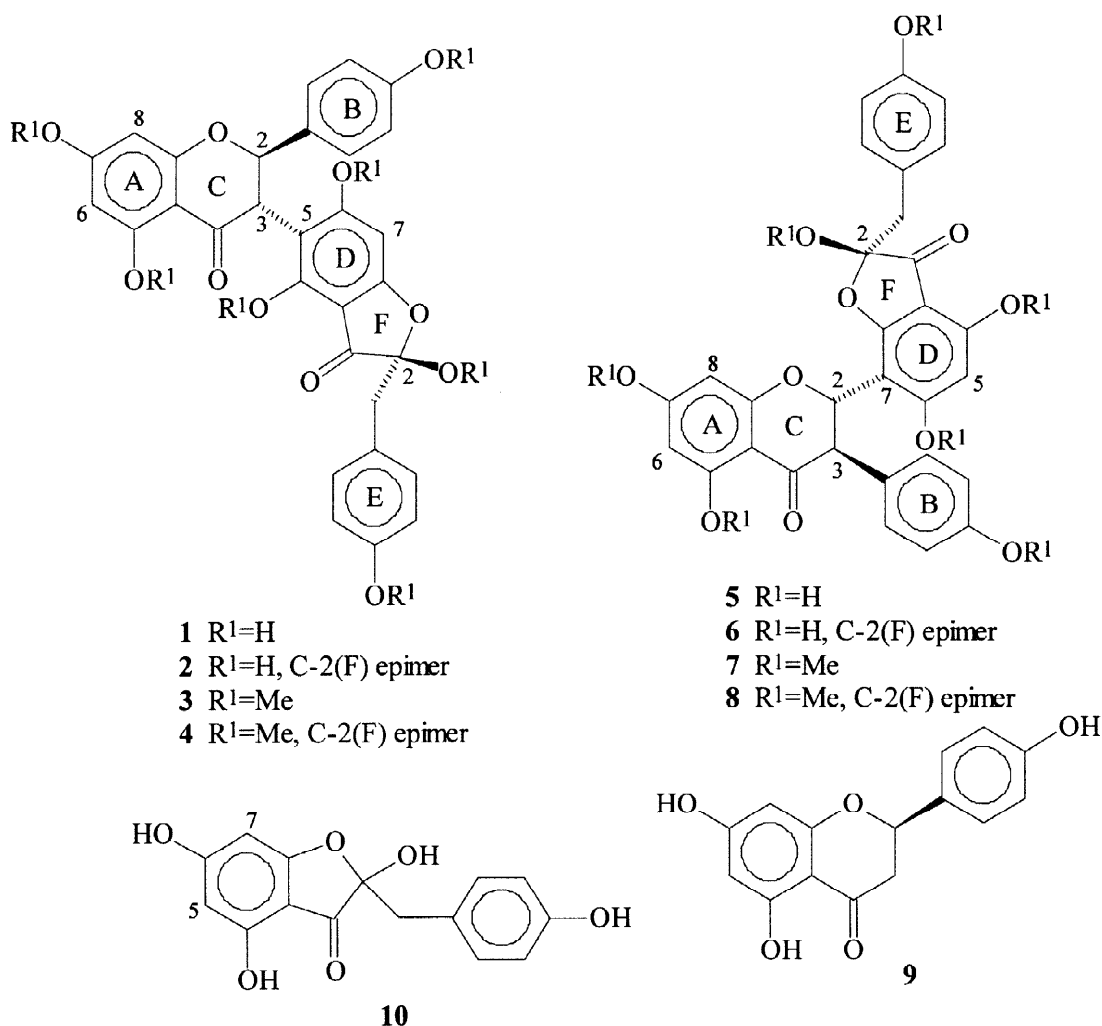
RESULTS AND DISCUSSION

The complex mixture of polyphenolic compounds¹⁻⁵ from the heartwood of *B. zeyheri* was extensively

* Part 3 is ref. 1, Part 2 is ref. 3 and Part 1 is ref. 2.

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fractionated and chromatographed into various fractions.^{1,5} These fractions still comprised mixtures and, following confirmation of the absence of natural methoxy groups by ¹H NMR, were methylated with dimethyl sulfate under rigorously anhydrous conditions. The additional chromatographic step offered by derivatization was a prerequisite for sample purity.



(2*R*,3*S*)-Naringenin-(3 α →5)-(2*R*)-maesopsin **1** and its 2-C(F) diastereomer **2** were purified as the yellow amorphous permethylaryl ethers **3** and **4**. Their ¹H NMR spectra exhibit the typical effects of slow dynamic rotational isomerism about the interflavonoid bond at ambient temperature. At elevated temperature (70°C), however, the close structural relationships with the naringenin-(3 α →7)-maesopsin diastereomers^{2,3} become obvious and the presence of a tetra-*O*-methylmaesopsin unit coupled with a tri-*O*-methylnaringenin moiety can clearly be discerned from the ¹H NMR data.

The ¹H NMR spectra of both diastereomers **3** and **4** exhibit the characteristic signals of the naringenin unit substituted at 3-C(C), resulting in an AM-spin system associated with 2-H(C) (δ 5.51, 5.65 for **3** and **4**, resp.) and 3-H(C) (δ 4.47, 4.56 for **3** and **4**, resp.); ³J_{2,3} = 12.0 Hz for both), an aromatic AB-system (J_{AB} = 2.0

Hz for both) representing the A-ring, and an AA'BB'-spinsystem ($J_{AB} = 9.0$ Hz for both) originating from the *para*-substituted B-ring. NOE associations between 2-H(C) and 2/6-H(B) (δ 7.02, 7.15 for **3** and **4**, resp.)

Table : ^1H NMR chemical shifts (δ_{H}) for biflavonoid derivatives **3**, **4**, **7** and **8**. Multiplicities and coupling constants (Hz) are given in parenthesis.

| Ring | Proton | 3 | 4 | 7 | 8 |
|------|--------------------|---|---|---|--|
| A | 6-H ^a | 6.16 (d, 2.0) | 6.15 (d, 2.0) | 5.99 (d, 2.0) | 6.05 (d, 2.0) |
| | 8-H ^a | 6.18 (d, 2.0) | 6.18 (d, 2.0) | 5.98 (d, 2.0) | 6.00 (d, 2.0) |
| B | 2,6-H | 7.02 (d, 9.0) | 7.15 (d, 9.0) | 7.32 (d, 8.5) | 7.38 (d, 8.5) |
| | 3,5-H ^b | 6.75 (d, 9.0) | 6.76 (d, 9.0) | 6.84 (d, 8.5) | 6.82 (d, 8.5) |
| C | 2-H | 5.51 (d, 12.0) | 5.65 (d, 12.0) | 5.61 (d, 10.5) | 5.62 (d, 10.5) |
| | 3-H | 4.47 (d, 12.0) | 4.56 (d, 12.0) | 4.66 (d, 10.5) | 4.62 (d, 10.5) |
| D | 5-H | | | 5.93 (s) | 5.95 (s) |
| | 7-H | 5.99 (s) | 6.08 (s) | | |
| E | 2,6-H | 7.13 (d, 9.0) | 7.11 (d, 9.0) | 7.01 (d, 8.5) | 7.15 (d, 8.5) |
| | 3,5-H ^b | 6.74 (d, 9.0) | 6.74 (d, 9.0) | 6.65 (d, 8.5) | 6.62 (d, 8.5) |
| F | 2-CH ₂ | 3.18 (d, 14.0) | 3.15 (d, 14.0) | 3.13 (d, 14.0) | 3.19 (d, 14.0) |
| | | 3.11 (d, 14.0) | 3.04 (d, 14.0) | 3.05 (d, 14.0) | 3.06 (d, 14.0) |
| | 2-OMe | 3.29 (s) | 3.21 (s) | 3.24 (s) | 3.19 (s) |
| | Ar-OMe | 3.93, 3.89, 3.84, 3.79, 3.76, 3.55 (each s) | 3.89, 3.84, 3.80, 3.79, 3.75, 3.69 (each s) | 3.89, 3.87, 3.86, 3.79, 3.78, 3.73 (each s) | 3.92, 3.88 (x2), 3.82, 3.79, 3.71 (each s) |

^a May be interchanged in **3**, **4**, **7** and **8** ; ^b May be interchanged in **3** and **4**

serve to identify the latter, while 2/6-H(B) and 3/5-H(B) (δ 6.75, 6.76 for **3** and **4**, resp.) may be associated by correlations in their COSY spectra. The close proximity, however, of 3/5-H(B) and 3/5-H(E) (δ 6.74, 6.74 for **3** and **4**, resp.) precludes the unambiguous differentiation of these protons for both compounds. NOE experiments distinguish 6- and 8-H(A) (δ 6.16, 6.18 and 6.15, 6.18 for **3** and **4**, resp.) by associations with 5- and/or 7-OMe(A). The maesopsin unit, substituted on the A-ring, is characterized by the conspicuously shielded 2-OMe(F) (δ 3.29, 3.21 for **3** and **4**, resp.) and the diastereotopic α -methylene (δ 3.18, 3.11 for **3** and δ 3.11, 3.04 for **4**, $J_{AB} = 14.0$ Hz for both) resonances reminiscent of the benzofuranoid moiety, together with an AA'BB'-spin system and a residual proton (δ 5.99, 6.08 for **3** and **4**, resp.). An NOE association between this proton and only one methoxy group, 6-OMe(D) (δ 3.55, 3.69 for **3** and **4**, resp.), define both compounds as 5(D)-linked dimers. NOE association of 2/6-H(E) (δ 7.13, 7.11 for **3** and **4**, resp.) with the respective α -methylene protons, permit access to the E-ring protons.

The close structural relationships of the permethylaryl ethers **7** and **8** with **3** and **4** are evident from the conspicuous resemblance of their ^1H NMR spectra (Table). The former pair, however, are notably free of the effects of dynamic rotational isomerism about the interflavonoid bond. An AM-spin system emanating from 2-H(C) (δ 5.61, 5.62 for **7** and **8**, resp.) and 3-H(C) (δ 4.66, 4.62 for **7** and **8**, resp.) once more discloses the presence of a heterocyclic C-ring with 2,3-*trans* relative configuration ($^3J_{2,3} = 10.5$ Hz for both) while an AB-

spin system ($J_{AB} = 2.0$ Hz for both) represents 6- and 8-H of the A-ring. Elements of a tetra-*O*-methylmaesopsin unit substituted at the A-ring are evident from the residual singlet (δ 5.93, 5.95 for **7** and **8**, resp.) and the presence of the shielded 2-OMe(F) (δ 3.24, 3.19 for **7** and **8**, resp.) together with the 2(F)-methylene ($J_{AB} = 14.0$ Hz for both). Differentiation between the two very similar AA'BB'-spin systems, reminiscent of the B- and E-ring protons, are effected *via* COSY spectra using the benzylic 2(F)-methylene and 3-H(C) resonances as reference signals.

Our initial communication¹ reported an NOE-association of the residual singlet with one methoxy group [6-OMe(D)] only, thus defining the compounds as 5(D)-linked dimers. Improved resolution of the NOESY data has since, however, revealed association of the residual singlet with two methoxy groups [4(D)- and 6-OMe(D) δ 3.89 and 3.87; 3.88 and 3.92, resp.] and we accordingly amend our initial structures to those of 7(D)-linked dimers **7** and **8**. Long-range COSY- and NOESY-experiments permit definition of compounds based on a 3-substituted flavanone by association of 2/6-H(B) with 2-H(C), as displayed by the 3 α (C) \rightarrow 5(D)-linked dimers **3** and **4**. This contrasts notably with the analogues **7** and **8** where coupling of 2/6-H(B) occurs with 3-H(C), indicative of a 2(C) \rightarrow 3(C)-'shift' of the B-ring and therefore of 2 α (C) \rightarrow 7(D)-linked dimers based on a 2-substituted isoflavanone moiety.

The above structural assignments correlate with the EI mass spectral fragmentation data of the derivatives **3**, **4**, **7** and **8**, which apart from the molecular ion M^+ , m/z 656, all display prominent peaks at m/z 535, 476, 355, 313, 312, 181, and 121. The m/z 121 fragment reflects the loss of a 4-methoxybenzyl radical involving the E-ring hence affording the m/z 535 ion, whereas that at m/z 476 results from the equivalent of an RDA-fragmentation of the ABC units of the flavanone- **3** and **4** or isoflavanone moieties **7** and **8**.

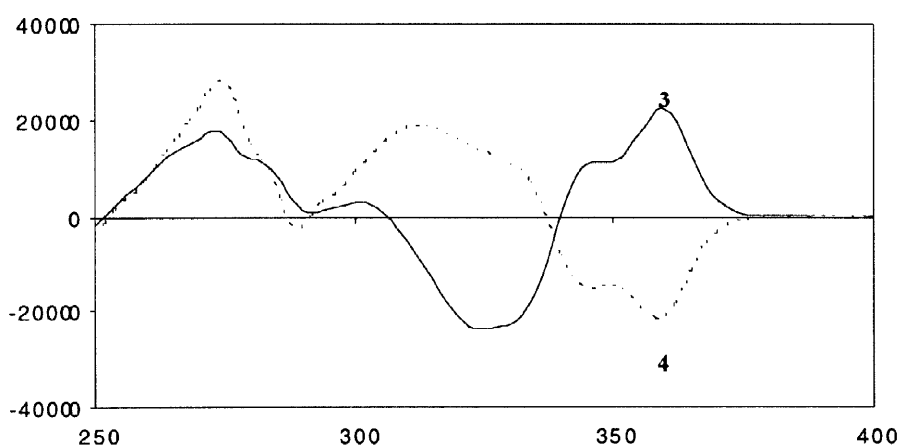


Figure 1 : CD spectra of biflavonoid derivatives **3** and **4**

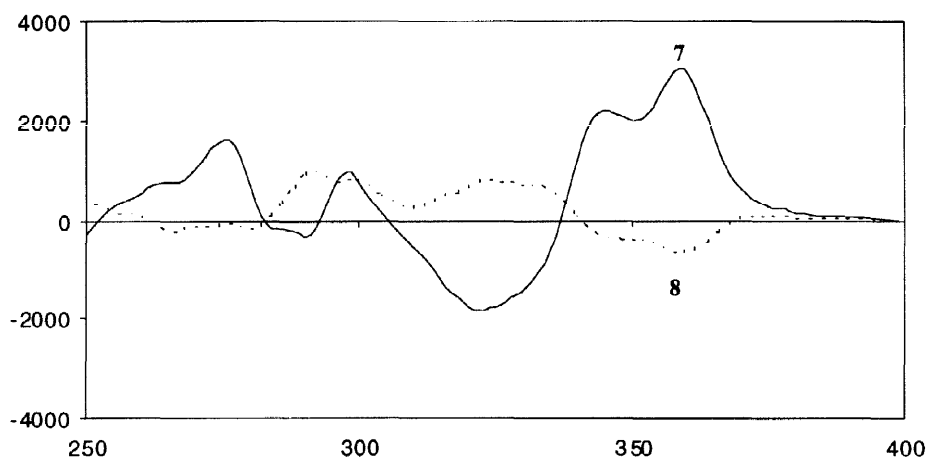


Figure 2 : CD spectra of biflavonoid derivatives **7** and **8**

The CD-curves (Figure 1) of the flavanone-(3 α →5)-benzofuranone analogues **3** and **4** are nearly superimposable with those of the (3 α →7)-coupled diastereomers.^{2,3} Thus, from the CD data and supporting evidence generated for the latter compounds it is clear that the positive Cotton effect observed for the $n \rightarrow \pi^*$ transition in the 340 - 380 nm region⁶ for **3** corresponds with a 2*R*(F) absolute configuration for the maesopsin unit. Similarly, the negative Cotton effect in the same region observed for **4** indicates a 2*S*(F) absolute configuration for the maesopsin moiety. In agreement with the chiroptical data of the flavanone-(3 α →7)-benzofuranone diastereomers,^{2,3} the positive Cotton effect observed for the $\pi \rightarrow \pi^*$ transition (260 - 290 nm) then indicates a 2*R*(C) absolute configuration for the flavanone unit⁷ in both **3** and **4**. When taken in conjunction with the 2,3-*trans* configuration of the heterocyclic C-ring indicated by ¹H NMR coupling constants, this corresponds with a 2*R*,3*S*(C) absolute configuration for the flavanone unit in both compounds and thus define them as (2*R*,3*S*)-naringenin-(3 α →5)-(2*R*)-maesopsin **1** and its 2*R*,3*S*(C):2*S*(F)-diastereomer **2**.

Although similar positive and negative Cotton effects in the 360 nm region⁶ $\{[\theta]_{359} +3.1 \times 10^3$ and $[\theta]_{360} -6.9 \times 10^2\}$ (Figure 2) facilitate the allocation of 2*R*(F)- and 2*S*(F)- configurations, respectively, to the isoflavonoid analogues **7** and **8**, Cotton effects originating from the isoflavanone moieties are not comparable with those of flavanones. Isoflavanones display a distinctive Cotton effect in the *ca.* 350 nm region for their $n \rightarrow \pi^*$ transitions, a positive effect reflecting a 3*R*-configuration and a negative effect a 3*S*-configuration.^{8,9} The strong positive effect observed in this region for compound **7** is therefore reminiscent of a 3*R*-configuration, the high amplitude being the result of the cumulative effect of the *R*-chirality at both 3-C(C) and 2-C(F). The much weaker and negative Cotton effect displayed by the isomer **8** would then likewise be in accord with a 3*R*(C)-configuration, since the positive effect originating from the chirality at 3-C(C) and the negative effect associated with a 2*S*(F)-configuration would now be opposing each other, the latter being slightly dominant (Figure 2). When taken in conjunction with the 2,3-*trans* relative configuration for the C-

ring protons ($^3J_{2,3} = 10.5$ Hz for both), a $2S(C)$ -configuration is implied for both compounds and therefore permits assignment of the absolute configuration of these novel biflavonoids as $(2S,3R)$ -dihydrogenistein- $(2\alpha\rightarrow7)$ - $(2R)$ -maesopsin **5** and its diastereomer, $(2S,3R)$ -dihydrogenistein- $(2\alpha\rightarrow7)$ - $(2S)$ -maesopsin **6**.

The absolute configuration of the isoflavanone ABC moieties $(2S,3R)$ in **5** and **6**, relative to those of the flavanone moieties $(2R,3S)$ in **3** and **4**, are in agreement with the proposed biogenetic origin of the biflavonoids **5** and **6** via initial 2,3-phenyl migration of the $(2R)$ -4',5,7-trihydroxyflavanone (naringenin) **9**, which coexists in *B. zeyheri*.⁵ Irrespective of the particular mechanism involved, *i.e.*, ionic or radical, several of which have been proposed,¹⁰⁻¹² such a migration always proceeds in a *suprafacial* fashion. $(2R)$ -Naringenin would therefore experience phenyl migration from C-2 β to C-3 β , leaving C-2 electron deficient and prone to nucleophilic attack from the less hindered α -face by the phloroglucinol-type D-ring of maesopsin **10**. Since the latter coexists as the racemate in *B. zeyheri*,⁵ the two diastereomers $(2S,3R)$ -dihydrogenistein- $(2\alpha\rightarrow7)$ - $(2R)$ -maesopsin **5** and $(2S,3R)$ -dihydrogenistein- $(2\alpha\rightarrow7)$ - $(2S)$ -maesopsin **6** originate. Proposals regarding the possible biogenetic origin of the flavanone analogues **1** and **2** are believed to parallel those of the zeyherin epimers which have been covered previously.³

EXPERIMENTAL

^1H NMR Spectra were recorded on a Bruker AM-300 spectrometer for solutions in CDCl_3 with Me_4Si as internal standard. Mass spectra were obtained with a Kratos MS-80 instrument, CD data in MeOH on a JASCO J-710 spectropolarimeter and IR spectra in CHCl_3 with a HITACHI 270-50 infrared spectro-photometer. TLC was performed on precoated Merck plastic sheets (silica gel 60 PF₂₅₄, 0.25 mm) and the plates were sprayed with H_2SO_4 -HCHO (40:1, v/v) after development. Preparative plates (PLC), 20x20 cm, Kieselgel PF₂₅₄ (1.0 mm) were air-dried and used without prior activation. Separations on Sephadex LH-20 were in EtOH at a flow rate of *ca.* 0.5 ml min⁻¹ (30 min fractions). Flash column chromatography (FCC) was carried out in a glass column (5 cm diameter) charged with Merck Kieselgel 60 (230-400 mesh) at a flow rate of 3 ml min⁻¹ (30 ml fractions) under N_2 pressure. Methylations were performed with Me_2SO_4 in dry Me_2CO containing anhydrous K_2CO_3 at reflux temperature. Water-soluble phenolics were freeze-dried with a Virtis Freeze mobile 12 SL. Evaporations were carried out under reduced pressure at *ca.* 40°C in a rotary evaporator.

The extraction of the heartwood of *B. zeyheri* with aqueous Me_2CO (8:2, v/v) and fractionation of the extract by means of countercurrent distribution and column chromatography using Sephadex LH-20 leading to fractions 7-8.9.1 to 7-8.9.25 were fully described in ref. 5. Methylation of fraction 7-8.9.15 (600 mg) and subsequent FCC in hexane-benzene- Me_2CO -MeOH (40:40:15:5, v/v) afforded three fractions 7-8.9.15.1 (tubes 1-18, 219 mg), 7-8.9.15.2 (19-28, 111 mg) and 7-8.9.15.3 (36-51, 183 mg). Fraction 7-8.9.15.1 was separated further (see ref. 5) to yield fraction 7-8.9.15.1.11 which was resolved into two components **3** (R_F 0.66) and **4** (R_F 0.62) by PLC in CHCl_3 -Et₂O (98:2, v/v, x4).

(2R,3S)-4',5,7-tri-O-methylnaringenin-(3 α →5)-(2R)-2,4,4',6-tetra-O-methylmaesopsin 3. The title compound (15 mg) was obtained as a *yellow amorphous solid*. (Found: M^+ , 656.2257. $C_{37}H_{36}O_{11}$ requires M , 656.2258); δ_H (Table); CD $[\theta]_{359.3}$ 2.2×10^4 , $[\theta]_{346.2}$ 1.2×10^4 , $[\theta]_{325.0}$ -2.4×10^4 , $[\theta]_{272.9}$ 1.8×10^4 (Figure 1); IR 1706, 1674 cm^{-1} .

(2R,3S)-4',5,7-tri-O-methylnaringenin-(3 α →5)-(2S)-2,4,4',6-tetra-O-methylmaesopsin 4. Compound 4 (13 mg) was obtained as a *yellow amorphous solid*. (Found: M^+ , 656.2257. $C_{37}H_{36}O_{11}$ requires M , 656,2258): δ_H (Table); CD $[\theta]_{358.6}$ -2.2×10^4 , $[\theta]_{345.4}$ -1.5×10^4 , $[\theta]_{312.4}$ 1.9×10^4 , $[\theta]_{274.4}$ 2.19×10^4 (Figure 1); IR 1708, 1680 cm^{-1} .

Fraction 6 (14.5 g) from the countercurrent distribution (see ref. 5) was separated by column chromatography using Sephadex LH-20 in EtOH to give the following fractions : 6.1 (tubes 1-38, 7.1 g), 6.2 (39-49, 560 mg), 6.3 (50-92, 856 mg), 6.4 (93-112, 134 mg), 6.5 (113-133, 202 mg), 6.6 (134-161, 67 mg) and 6.7 (162-347, 148 mg). Fraction 6.4 (134 mg) was methylated and purified by PLC in benzene-Me₂CO (7:1, v/v) to yield a mixture (R_F 0.24) which was resolved into two components 7 (R_F 0.63) and 8 (R_F 0.53) by a further PLC separation in benzene-EtOAc (3:1, v/v, x7).

(2S,3R)-4',5,7-tri-O-methyldihydrogenistein-(2 α →7)-(2R)-2,4,4',6-tetra-O-methylmaesopsin 7. The title compound (5 mg) was obtained as a *yellow amorphous solid*. (Found: M^+ , 565.2257. $C_{37}H_{36}O_{11}$ requires M , 565.2258); δ_H (Table); CD $[\theta]_{359.0}$ 3.1×10^3 , $[\theta]_{345.4}$ 2.2×10^3 , $[\theta]_{322.1}$ -1.8×10^3 , $[\theta]_{297.7}$ 9.8×10^2 , $[\theta]_{290.0}$ -3.73×10^2 , $[\theta]_{275.2}$ 1.6×10^3 (Figure 2); IR 1706, 1702 cm^{-1} .

(2S,3R)-4',5,7-tri-O-methyldihydrogenistein-(2 α →7)-(2S)-2,4,4',6-tetra-O-methylmaesopsin 8. Compound 8 (6 mg) was obtained as a *yellow amorphous solid*. (Found: M^+ , 565.2257. $C_{37}H_{36}O_{11}$ requires M , 565.2258); δ_H (Table); CD $[\theta]_{359.7}$ -6.9×10^2 , $[\theta]_{349.4}$ -3.8×10^2 , $[\theta]_{324.5}$ 8.0×10^2 , $[\theta]_{310.0}$ 2.6×10^2 , $[\theta]_{299.1}$ 8.5×10^2 , $[\theta]_{291.4}$ 1.0×10^3 (Figure 2); IR 1707, 1701 cm^{-1} .

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